



© VMS RESEARCH FOUNDATION

ANTIMICROBIAL POTENTIAL AND GC-MS ANALYSIS OF BRYOPHYTE TARGIONIA HYPOPHYLA L.

Wankhede T.B.

Department of Botany, Shri Shivaji Science College, Amravati- 444603, M.S. India Email: tusharwan@gmail.com

Communicated: 16.12.18

Accepted : 17.01.19

Published: 30.01.19

ABSTRACT:

Targionia hypophylla L., a terricolous liverwort occurs in close vicinity of water identified with carnation of violet blue or black ventral sporophytes often seen during rainy seasons. Thalli light to dark green, in close clusters, overlapping, usually with apical end projecting outward and downward. Thallus thin, somewhat brittle, abovate to linear - oblong, simple or once, rarely dichotomously branched, 10-15 mm. long and 3-6 mm. wide. Present exploration carried out to trace the antimicrobial potential of the bryophytic division of plants having pioneer status during conquest of land. At first instance the preliminary phytochemical characteristics were analysed. The plant extract in different solvents like ethanol, methanol, chloroform, acetone and petroleum ether and water were tested against different gram negative and gram positive bacterial and fungal strains for antimicrobial sensitivity test. Further, using sophisticated techniques like GC-MS chemical analysis of plant carried out to trace the potential chemical content with respect to future elucidation of compounds for pharmacology.

Key words :- Liverwort, Antimicrobial test, GC MS-Analysis

INTRODUCTION:

Bryophytes are the most fascinating cryptogramic plants occurs in nature with close vicinity of water bodies which dwells first on the land. Bryophytes are more common in humid areas and during rainy seasons, but usually show a preference for microclimatic niches such as crevices of rocks and trees and the vicinity of small shady springs (Shaw and Renzaglia, 2004). Bryophytes make a significant contribution to the floral diversity of this "watery planet" and since its inception constitute an important component of the forest ecosystem being the first colonizers on variety of habitats. They are highly specific group of plants with about 25,000 species distributed the world over, making it the second largest group of land plants after angiosperms (Alam et al., 2011). The occurrence of antimicrobial substances in thalli of bryophytes has been reported by Madsen and Pates (1952) for the first time in bryophyte Sphagnum portoricense and Sphagnum strictum against pathogens Staphylococcus aureus and Pseudomonas aeruginosa. Wolter (1964) screened antifungal activity of 18 species of bryophytes belonging to Pellia epiphylla and Diplophyllum albicans and 16 other mosses. Banerjee and Sen (1979) by examining 52 species (40 genera) of the bryophyte where 29 species showed activity against at least one bacterium. The liverworts Asterella sanguinea, Marchantia paleacea and the moss Brachythecium procumbens showed the broadest spectrum of antibacterial activity. Latiff et al., (1989) screened 14 moss species belonging to 10 families of Malaysia for antibacterial activity.

The ethanolic extract of the mosses were tested against E.coli, S. aureus and Bacillus subtilis. S. aureus was the most susceptible bacterium which was inhibited by all the moss extract. Grammes et al., (1994) carried out in-vitro culture of Fossombronia pusilla and isolated or analyzed terpenoids, as same produced in natural condition of plants. Mewari and Kumar (2008) used crude methanol and flavonoid extracts of Marchantia polymorpha L. against three bacterial strain viz., E. coli, P. mirabilis, S. aureus and four fungal strains viz., A. flavus, A. niger, C. albicans and T. mentagrophytes for antimicrobial screening. Sawant and Karagade, (2010) investigated in vitro antimicrobial activity extracts of three liverworts, T. hypophylla, P. intermedium, and A. wallichiana in various solvents have a broad spectrum of activity. Krishnan et al., (2012) reported in-vitro microbicidal potentiality of Targionia hypophylla L. and Bryum species of bryophytes. Asakawa (2001) reported the chemical constituents from the bryophytes like acetogenins, monoterpenoids, sesquiterpenoids, diterpenoids, triterpenoids and bis (bibenzyl)s from Japanease, Taiwanese, New Zealand, Argentinean and European regions. Ücüncü et al., (2010) analyzed the chemical constituents of mosses like Torula, Homalothecium, Hypnum and Pohila species using GC-MS techniques and recorded presence of many essential oil with antimicrobial activity. Russell (2010) reported that, the extracts of some bryophytes in South Western British Columbia, possess novel chemical compounds with antimicrobial activity.

MATERIAL & METHODS:

The thalli of Targionia hypophyla were collected from Melghat forest of Amravati District and cleaned carefully and washed under tap water followed by shade drying to use in powder form. Using Soxhlet apparatus, the powdered samples of plants were extracted in ethanol, methanol, petroleum ether, chloroform and acetone and different solvent and evaporated by rotary evaporator in controlled condition of temperature to get dried extracts were stored in labelled sterile wide mouthed screw capped bottles at 4°C and used for further study (Parekh and Chanda, 2008). The standard pathogenic bacterial and fungal strain cultures were procured from Microbial Type Culture Collection and Gene Bank (IMTECH), Chandigarh, India. Disc diffusion method was used for the antibacterial sensitivity test by following the standard methods (NCCLS, 1990). Phytochemical analysis was done for probable phytoconstituents like alkaloids, flavonoids, tannins, phenolics, steroids, saponins and terpenoids regarding the nature of constituents present in crude drug (Sadashivam and Manickam, 2005). Methanol extract of the plant send for GC-MS analysis to Shivaji University, Kolhapur and results were obtained. GC-MS analysis of the sample was carried out using Shimadzu Make QP-2010 with non-polar 60 M RTX 5MS Column.

RESULT & DISCUSSION:

Antimicrobial sensitivity test of Targionia hypophylla

Plant extracts of liverwort T. hypophylla were obtained in different solvents were tested against test microorganism showed significant activity of antibiosis (Table-1). The aqueous extract of the plants were inhibitory against E. coli, S. aureus and fungus A. niger. However, the petroleum ether extracts showed no any response to the all microorganisms and reactions were nullified. The ethanol extracts were more effective with broad spectrum of antibiosis against most of the microorganisms except the S. flexneri and R. oryzae. The chloroform extracts of the plant showed positive result against six microorganisms i.e. E. coli, K. pneumoniae, S. flexneri, S. typhimurium, A. niger and C. albicans. However, the methanol extracts of the plant was found positive against five selected microorganisms like E. coli, K. pneumoniae, S. flexneri, S. typhimurium, and C. albicans. The acetone extracts showed very promising response of action in all the extracts except K. pneumoniae and R. oryzae. Among all the extracts, the petroleum ether extract was found

non-reactive against the entire microorganism. However, slight induction was observed in some petri plates rarely but non-recordable. The aqueous extract is less reactive than the other extracts but ethanol, chloroform, methanol and acetone extracts were more responsive to most of the pathogens. The maximum responses to all the extracts were found in microorganism *E. coli, S. typhimurium, A. niger* and *C. albicans* and less in *K. pneumoniae, S. flexneri*, and *S. aureus* or very less in *P. vulgaris*. Most of the extracts bear light green, yellow green, dark green and blackish green colour in crude form (Fig-1).

Phytochemical GC-MS analysis of T. hypophyla Most of the bryophytes are being used as medicinal plants in China, Europe, North America and rest of the world. Bryophytes have been applied as decoctions to cure various kinds of diseases. The present study emphasizes to elicit out probable phyto-constituents from the thalli of Targionia hypophylla experimental bryophytes which subjected for the phytochemical and GC-MS analysis. Preliminary phytochemical test showed the presence of alkaloids, flavonoids, sterols, tannins and terpenoids. Methanol extract was selected due to its antimicrobial results which were obtained previously. The liverwort Targionia hypophylla in GC MS analysis analysed on the basis of retention time, percent area of the peak and compound determination with molecular formula and molecular weight (Fig-2). On the basis of data received and GC-MS chromatogram, it showed the presence of different and diversified compounds like, Longifolene, Bicyclo [5.3.0] decane, 2-methylene-5-(1-methylvinyl)-8-methyl, Patchouli alcohol, n-Hexadecanoic acid and 9-Octadecenoic acid (Table -2).

Krishnan *et al.*, (2012) confirmed that, the bryophytes having alkaloids are pharmacologically active as they have physiological effects on human as well as other animals and serves as therapeutic and anti-malarial drugs. Petroleum ether extract of liverwort *Targionia hypophylla* were active against both gram positive and gram negative bacteria and fungi. *Targionia* were particularly active against bacteria *E. coli* and fungus *C. albicans* and *A. niger*. However, these species were less reactive to the bacteria *S. flexneri* and almost non-reactive to the fungus *R. oryzae*. These findings are in contrast with the findings of Banerjee and Sen (1979) that these liverworts are active against bacteria *P. aeruginosa*.

This may be due to the variations in chemical composition of particular species of plants, which can also vary according to the geographical origin and harvesting seasons (Burt, 2004). It also showed that specific antibacterial compounds, effective against the selected bacterial or fungal species, tends to be isolated more effectively from liverworts using ethanol and methanol.

The bryophyte Targionia possess compound Bicyclo (5.3.0)decane, 2-methylene-5-(1methylvinyl)-8-methyl. Chen et al., (2013) found this compound as potent composition with potential of anticancer activities among essential oil obtained from Myrrh and Frankincense. 9octadecenoic acid and 9, 12 octadecadienoic acid (z-2) methyl esters are commonly called Linoleic acid which is a polyunsaturated fatty acid commonly produced in plants. The bryophytes species Targionia showed occurrence of these compounds. Park et al., (2013) reported antimicrobial activity of gamma linolenic acid from Enteromorpha linza against several bacteria like Streptococcus mutans and fungi Candida albicans. The liverwort Targionia hypophylla showed the presence of chemical constituent Longifolene i.e. a tricyclic sesquiterpenoids. It commonly occurs in plants like pines, angiosperms, small amount in fungi and bryophytes. Ücüncü et al., (2010) reported antimicrobial activity of longifolene among various mosses like Torula, Hyphum and Pohila against bacteria E. coli, P. aeruginosa, S. aureus and fungi Candida albicans. This finding corresponds to the present investigations of antimicrobial activity of Targionia species. Hata et al. (1993) discovered an "Acne-Vulgaris" treatment comprising of longifolene, caryophyllene, centrene and thujopsene as a key ingredients and got U.S. patent no. US-S200429A for such a novel medicine. The compound Bicyclo (5.3.0) decane, 2methylene-5-(1-methylvinyl)-8-methyl. Chen et al., (2013) found this compound as potent composition with potential of anticancer activities among essential oil obtained from Myrrh and Frankincense.

Patchouli alcohol or patchouli is a terpene compound with typical patchouli scent and used in chemotherapy drugs. Yang *et al.*, (2013) evaluated the antimicrobial activity of patchouli alcohol and pogostone against various bacteria like *E. coli*, *P. aeruginosa*, *S .aureus* and *S. dysentyeriae*. The liverworts *Targionia*, *Asterella* and *Plagiochasma* spp. in present study found reactive to these bacterial pathogens with similar promising results.

9-octadecenoic acid and 9, 12 octadecadienoic acid (z-2) methyl esters are commonly called Linoleic acid which is a polyunsaturated fatty acid commonly produced in plants. The bryophytes *Targionia*, showed occurrence of these compounds. Park *et al.*, (2013) reported antimicrobial activity of gamma linolenic acid from *Enteromorpha linza* against several bacteria like *Streptococcus mutans* and fungi *Candida albicans*.

CONCLUSION:

Hence, in present investigation, many chemical compounds distributed variably and dominantly present among *Targionia* species showing corelation between chemical compound analyzed and their probable antimicrobial sensitivity against microorganisms. Hence, all the solvent extracts possesses some novel chemical compounds with highly antimicrobial potential is confirmed in the present investigation. Future studies on character elucidation of compounds will be remarkable insight for drug designing and benefits of mankind.

REFERENCES:-

- Alam, A., Sharma, V., Sharma, S.C. (2011) Bryoflora of the Ranthambore Tiger Reserve, Rajastan (India). *The Archives for Bryology* 106: 1-12.
- Asakawa, Y. (2001) Recent advances in Phytochemistry of bryophytes-Acetogenins, terpenoids and bis (benzyls) from selected Japanease, Taiwanese, New Zealand, Argentinean and European liverworts. *Phytochemistry*. 56: 297-312.
- Banerjee, R.D. and Sen, S.P. (1979) Antibiotic activity of the Bryophytes. American

Bryological and Lichenological Society. *The Bryologist.* 82 (2): 141-153.

- Burt, S. (2004) Essential Oils: their antibacterial properties and potential applications in foods - a review. *International Journal of Food Microbiology*. 94: 223-253
- Chen, Y., Zhou, C., Ge, Z., Liu, Y., Lui, Y., Feng, W., Li, S., Cheng, G., Wei, T. (2013) Compaction and potential anticancer activities of essential oil obtained from Myrrh and Frankincense: Oncology letters. 6 (4): 1140-1146.
- Dr. Duke's Phytochemical and Ethnobotanical Databasesttps://hytochem.nal.usda.gov/p hytochem.
- Grammes, C., Burkhardt, G. and Vecker, H. (1994) Triterpenes from *Fossombronia* liverworts. *Phytochemistry* 35: 1293-1296.
- Hata, H., Ishida K, Sato T, Tsukada S (1993) An Acne Vulgaris treating: US Patent No. US S200429A. (USA) :11-19.

 $_{\rm Page}79$

- Krishnan, R., Manaj, G.S. and Murugan K. (2012) In vitro microcidal potentiality of Targionia hypophylla L. and Bryum species-Bryophyte. International Journal of Pharmacy and Pharmaceutical Sciences 4 (2): 410-413.
- Latiff, A., Tumin, S.Z. and Mohamad, A.D.H. (1998) The effect of moss extracts on the growth of three species of bacteria. *Malaysian Applied Biology* (Malaysia). 18 (1): 77-84.
- Madsen, G.C. and Pates, A.L. (1952) Occurrence of antimicrobial substances in chlorophyllose plants growing in Florida. *Bot. Gaz.* 113: 203-300.
- Mewari, N. and Kumar, P. (2008) Antimicrobial activity of extracts of *Marchantia* polymorpha. Pharmaceutical Biology. 46 (10-11): 819-822.
- NCCLS (1990) Manual on "Performance Standards for Antimicrobial Disk Susceptibility Tests". Approved Standard NCCLS Publication, M2-A4, Villanova, PA, USA., (1990 a-b).
- Parekh, J. and Chanda, S. (2008) Antibacterial activity of aqueous and alcoholic extracts of 34 Indian Medicinal plants against some bacterial species. *Turk. J. Biol*, 32: 63-71.
- Park, N.H., Choi, J.S., Hwang, S.Y., Kim, Y.C. and Hang, Y.K. (2013) Antimicrobial activity of gamma linolenic acid from *Enteromorpha linza* against several bacteria. *Botanical Studies.* 34: 39-45.

- Russell, M.D. (2010) Antibiotic activity of extracts from some bryophytes in South Western British Columbia. MSJA. 2: 9-14.
- Sadashivam, S. and Manickam, A. (1996) Biochemical Methods, New Age International Publishers, New Delhi: 193-199.
- Sawant Ulka J. and Karadge B.A. (2010) Antimicrobial activity of some bryophytes (liverworts and hornwort) from Kolhapur District. *Pharmacognosy Journal*, 2 (16): 29-32
- Shaw, J. and Renzaglia L. (2004) Phylogeny and diversification of Bryophytes. American Journal of Botany. 91 (10): 1557-1581.
- Ücüncü, O., Cansu, T.B., Ozdemir, T., Koraglu, S.A., Yayli, N. (2010) Chemical composition and antimicrobial activity of essential oil of mosses *Torula muralis*. Hedw., *Homalothecium lutescence* Hedw., *Hypnum curpresstarm* Hedw. and *Pohila nutans* Hedw. from Turkey. *Turk J. Chem.* 34: 825-834.
- Wolters, B. (1964) Die Verbrietung antifungaler Eigenschaften bei Mossen. Planta. 62: 88-96.
- Yang, X., Zhang, X., Yang, S.P., Liu, W.Q. (2013) Evaluation of the antibacterial activity of patchouli oil. *Iranian Journal of Pharmaceutical Research.* 12 (3) : 307-316.

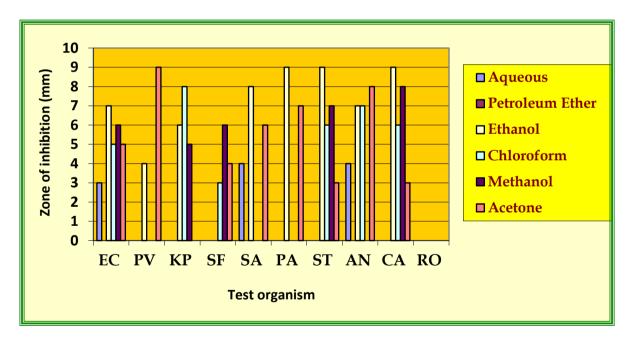
Plant	Solvent Extract	Zone of Inhibition [mm]								¥		
Herbal Preparation		EC	PV	KP	SF	SA	PA	ST	AN	CA	RO	AVERA GE
	Aqueous	3	0	0	0	4	0	0	4	0	0	
yla	Petroleum Ether	0	0	0	0	0	0	0	0	0	0	
ydo	Ethanol	07	04	06	0	08	09	09	07	09	0	
hypophyla	Chloroform	05	0	08	03	0	0	06	07	06	0	
	Methanol	06	0	05	06	0	0	07	0	08	0	
jion	Acetone	05	09	0	04	06	07	03	08	03	0	
Targionia	Ampicillin	22	20	27	29	34	31	30	-	-	-	
4	Nystatin	-	-	-	-	-	-	-	31	29	28	

Table- 1 : Antimicrobial sensitivity test of Targionia hypophylla

* Data represented in mean of three replicates.

*EC = Escherichia coli [MTCC-729], PV= Proteus vulgaris [MTCC-744], KP = Klebsiella pneumoniae [MTCC-661], SF = Shigella flexneri [MTCC-1457], SA= Staphylococcus aureus [MTCC-96], PA= Pseudomonas aeruginosa [MTCC-424], ST = Salmonella typhimurium [MTCC-98], AN = Aspergillus niger [MTCC-281], CA= Candida albicans [MTCC-227], RO= Rhizopus oryzae [MTCC-554]

Fig-2: Analysis of antimicrobial sensitivity of Targionia hypophylla



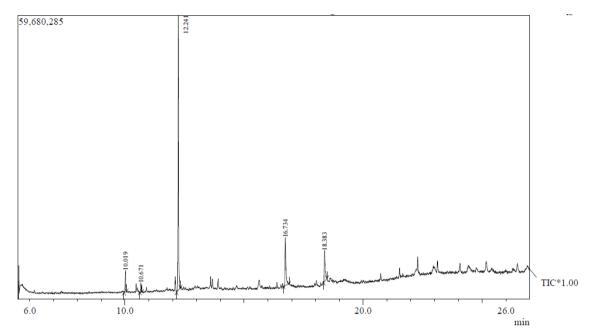


Fig 2: GC-MS chromatogram of Targionia hypophylla

Table : 2 GC-MS	analysis of	Taraionia	hupophulla
14010 . 2 40-110	analy 515 01	Iurgioniu	ngpopnynu

S r. N o.	Retent ion time	% area of the peak	Compound analyzed	Molecu lar formula	Molecu lar weight	Common Name	Activity reported
1	10.01	4.95	Longifolene	C15H24	204	Sesquiterpe ne	Acne vulgaris treatment, Antibacterial and Antifungal
2	10.66	2.17	Bicyclo[5.3.0]de cane, 2- methylene-5-(1- methylvinyl)-8- methyl	C15H24	204	Alkene	Anticancerous
3	12.24	65.25	Patchouli alcohol	C ₁₅ H ₂₆ O	222	Terpene	Antibacterial
4	16.73	16.37	n-Hexadecanoic acid	C ₁₆ H ₃₂ O ₂	256	Palmitic acid	Antioxidant, Hypocholesterole mic Nematicide, Pesticide
5	18.38	11.26	9-Octadecenoic acid	C ₁₈ H ₃₄ O ₂	282	Oleic acid	Antitumor and antibacterial activity.

(*Activity as per Data of Duke's library)